

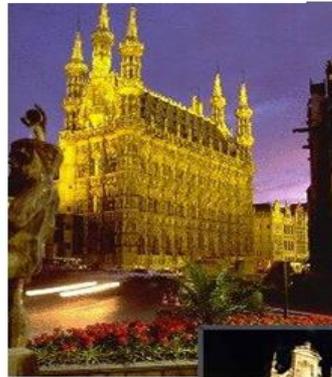
# Zika virus mouse model

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[www.antivirals.be](http://www.antivirals.be)

**KU LEUVEN**



WHO Zika Research  
Meeting, Geneva March  
3-5<sup>th</sup> 2016



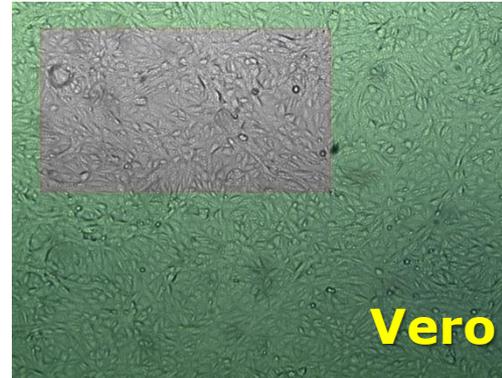
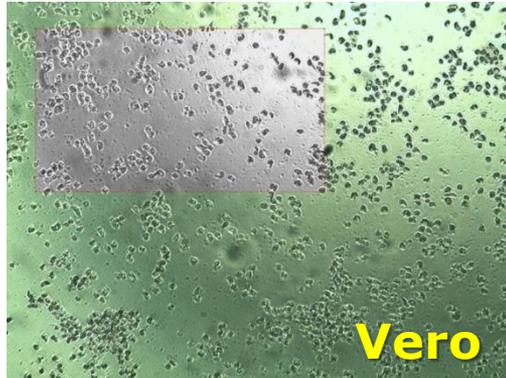
The Rega Instituut of the KU Leuven, the Wellcome Trust and Janssen are together joining the fight against Dengue fever (breakbone fever). In August 2013 they concluded a collaboration agreement for the development of a treatment or preventive medicine against Dengue fever virus infection.

The Dengue virus is potentially life-threatening. It is a virus spread by mosquitoes that infects hundreds of millions of people every year, chiefly in the developing countries. At present there is still no approved vaccine or specific treatment for the disease. The Rega Instituut and Janssen are pooling their expertise and resources in a joint research program. The Wellcome Trust will finance Rega Instituut's research activities. Janssen will obtain the possibility of taking out an exclusive, worldwide license for the development and commercialization of compounds that could be discovered within the context of this research program.

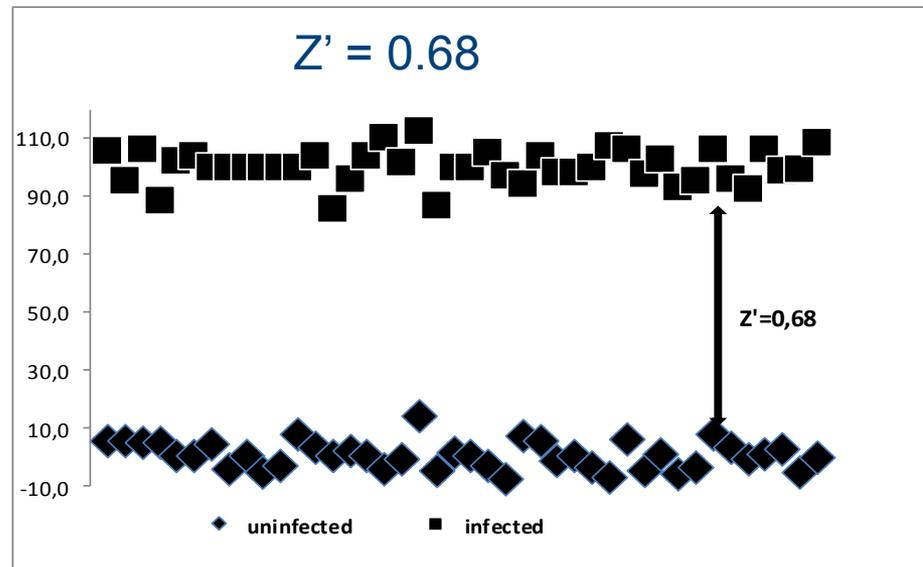
This collaboration is another example of Janssen's efforts to combat contagious diseases with a still largely unanswered medical need.



# ZIKV induced cytopathic effect

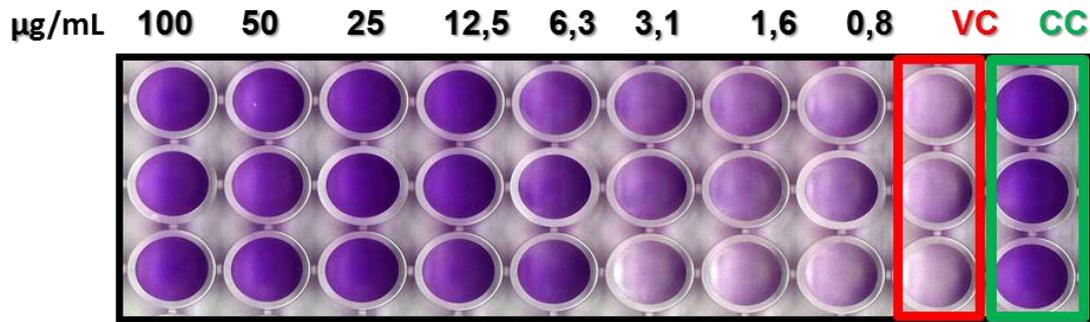


Strain MR766

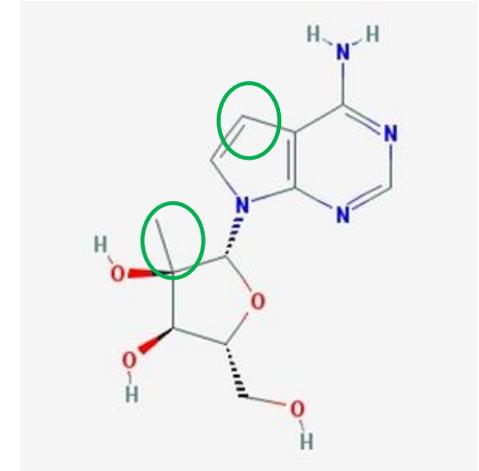


# Antiviral activity of 7DMA, *in vitro*

## Antiviral assay

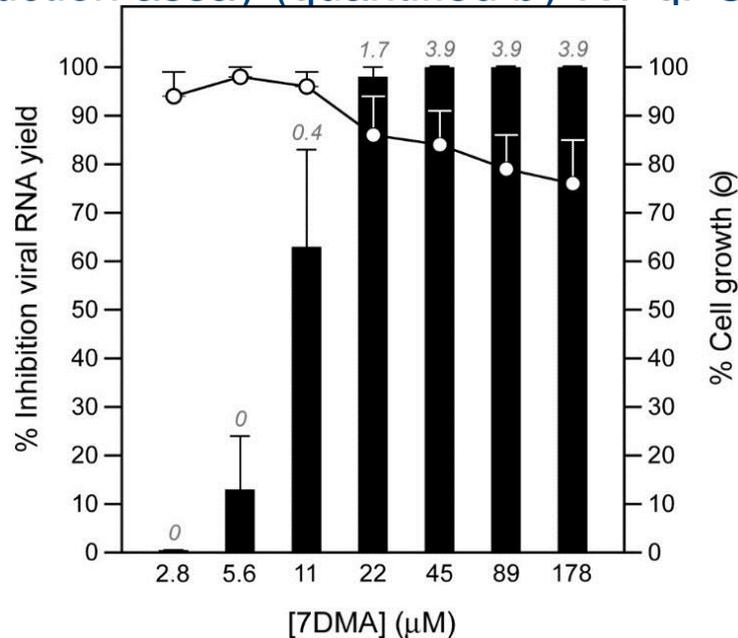


Also established for 384 well plate format



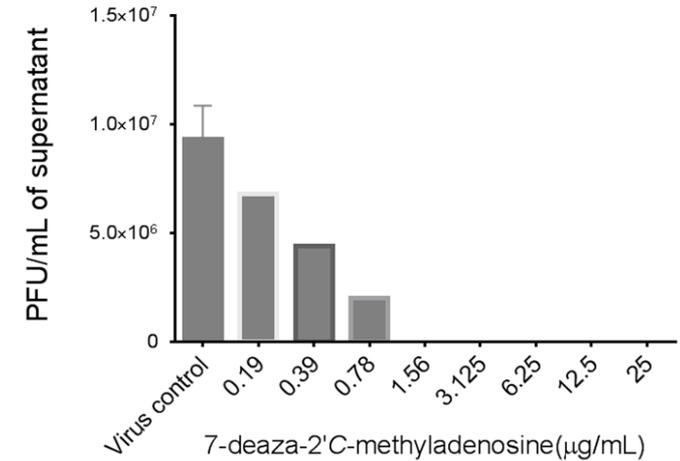
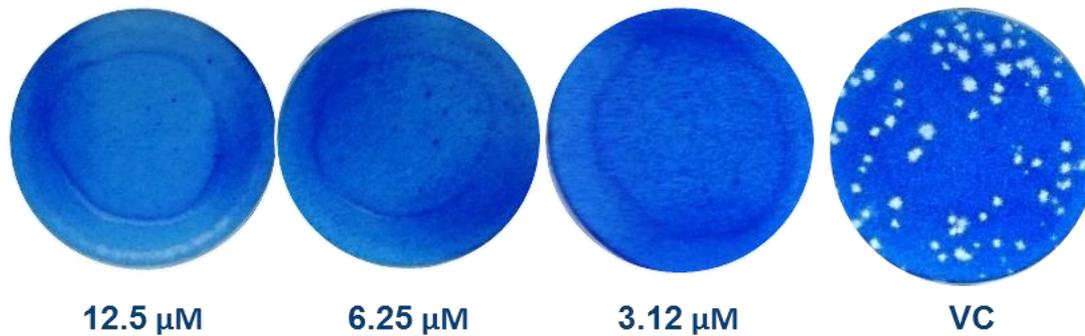
7-deaza-2'-C-methyladenosine

## Virus yield reduction assay (quantified by RT-qPCR)

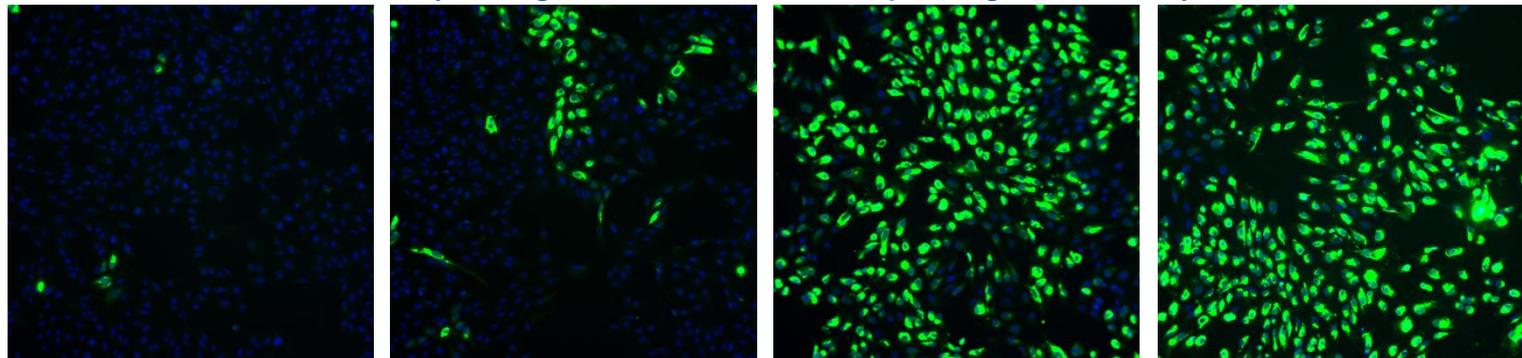


- $EC_{50} = 9.6 \pm 2.2 \mu M$

# Plaque reduction & viral antigen expression



Immunofluorescence assay using anti-Flavivirus Group Antigen Antibody, clone D1-4G2-4-15

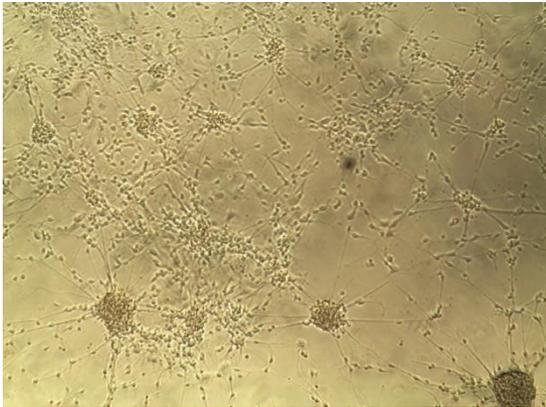


$$EC_{50} = 5.7 \pm 2.2 \mu\text{M}$$

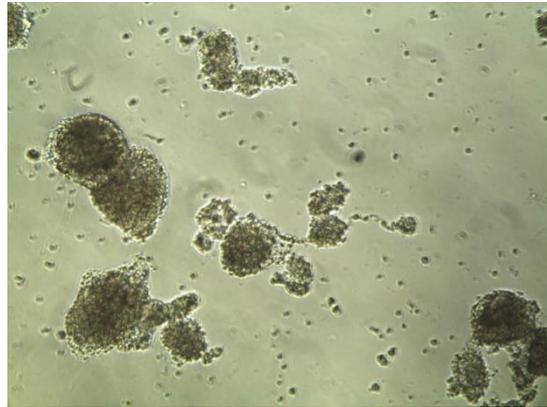
# ZIKV infection in stem-cell derived immature cortical neurons

Induced pluripotent stem cell (iPSC)-derived cortical neuron cells (immature) infected with Zika strain MR766 (MOI 0.1)

**uninfected**



**Infected (day 6 p.i.)**

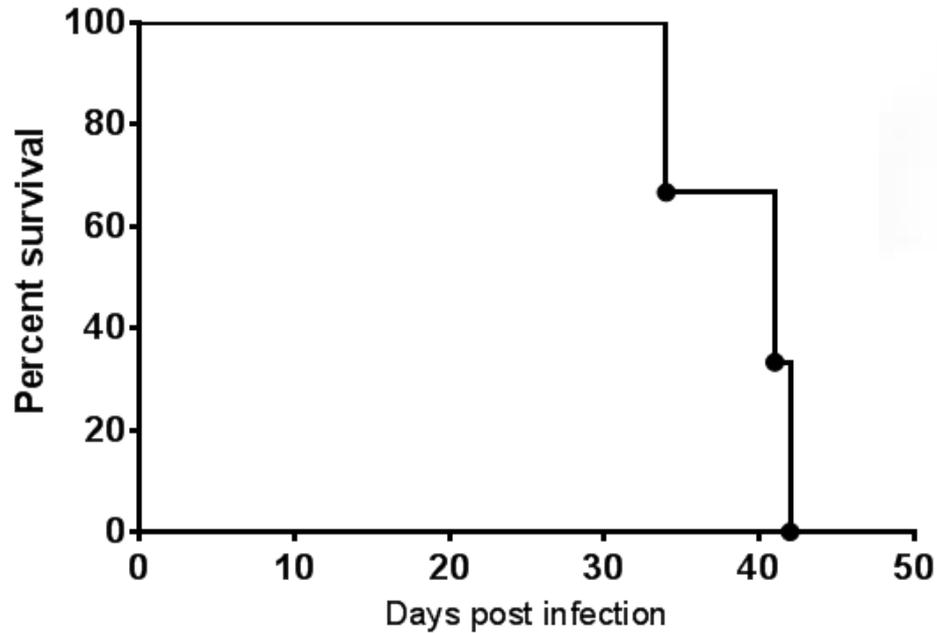


**Infected + 7DMA (day 6)**



# Infection of SCID mice with ZIKV

*pilot studies*

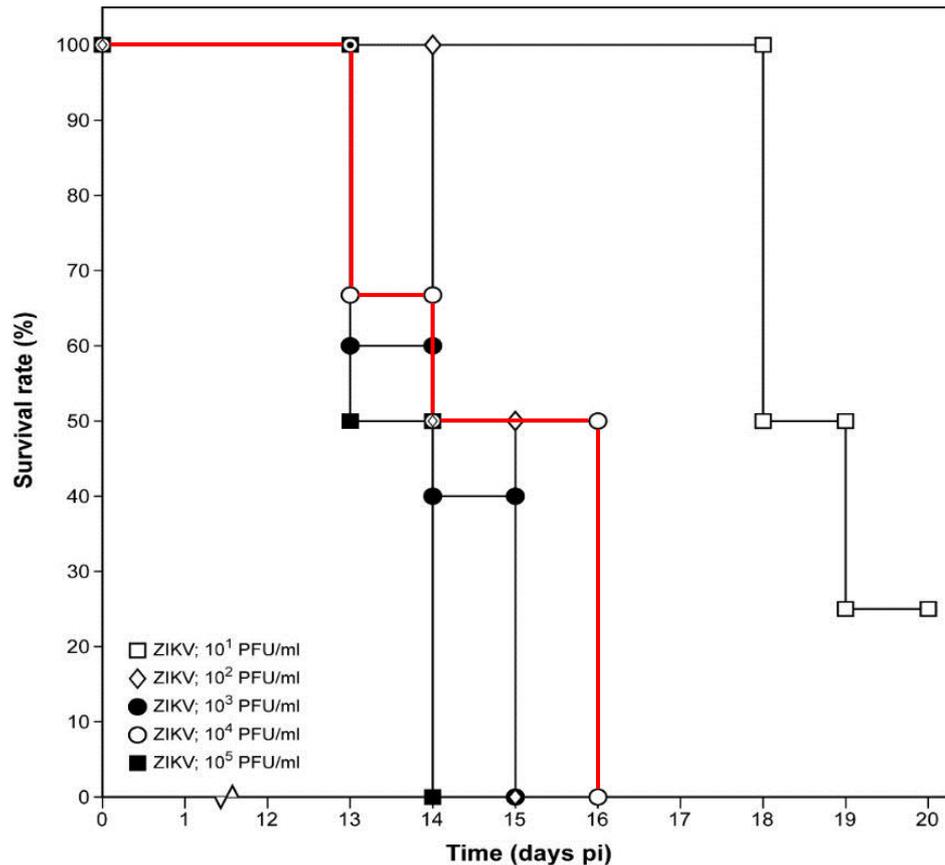


$10^4$  PFU/mL (200  $\mu$ L), ip

MDE =  $40.0 \pm 4.4$  d pi

# Infection of AG129 mice with ZIKV

*pilot studies*



IFN alfa, beta,  
gamma receptor KO

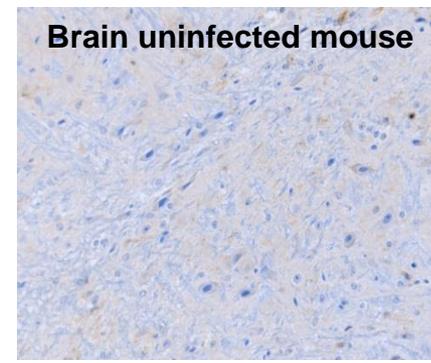
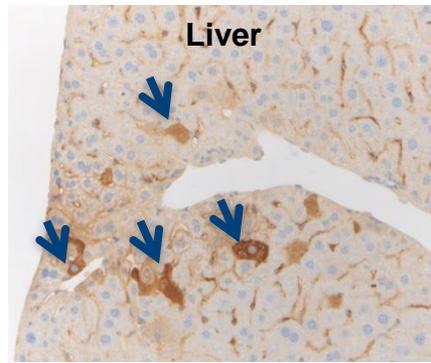
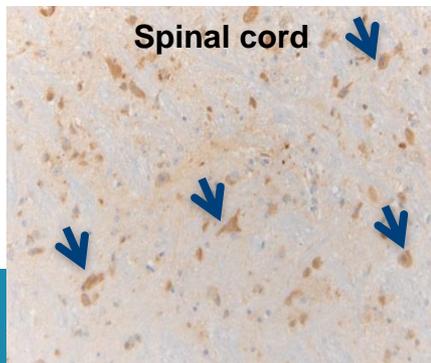
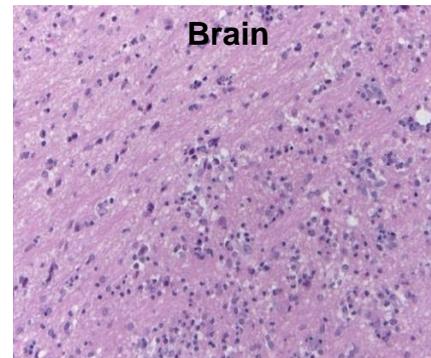
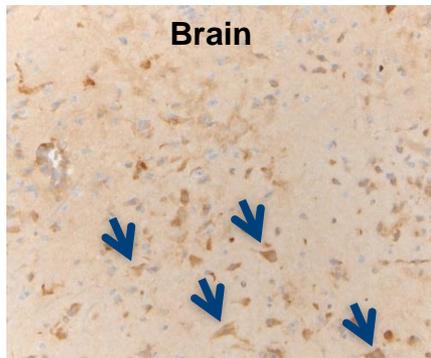
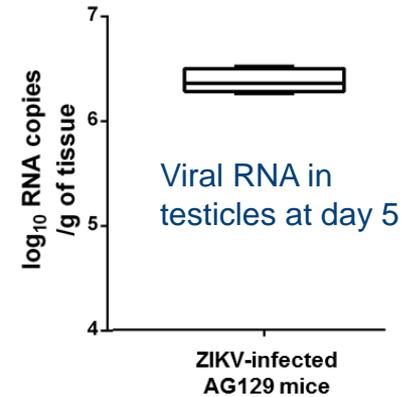
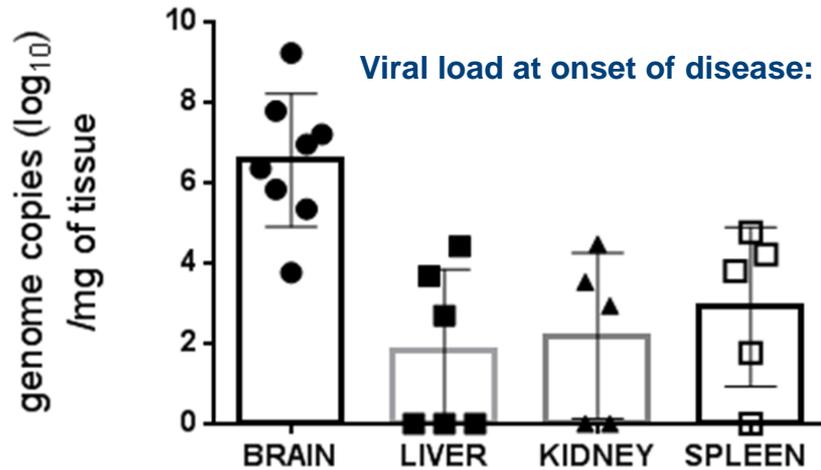
**$10^4$  PFU/mL (200  $\mu$ L), ip**

**MDE =  $15.0 \pm 1.5$  d pi**

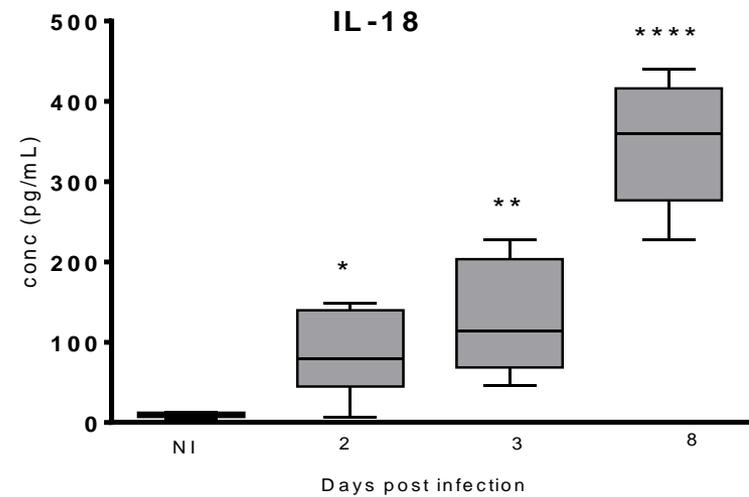
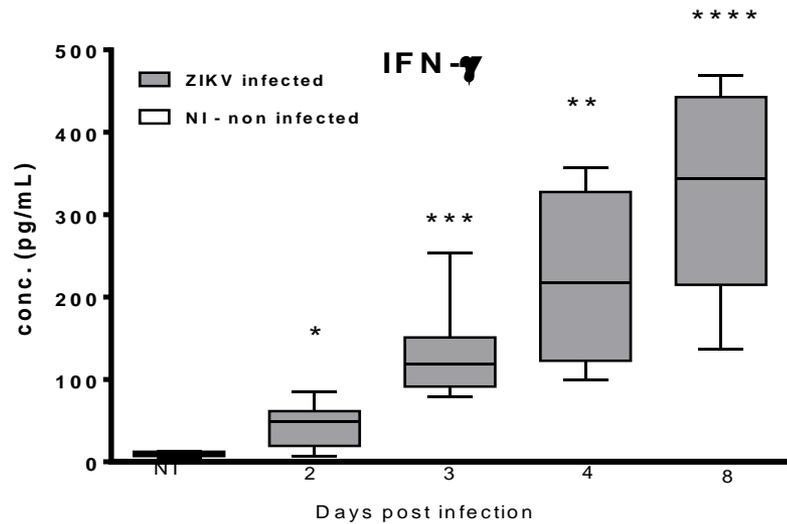
Symptoms	Days post infection	Affected/Total
paralysis of lower limbs	12-16	18/18
weight loss	13-16	18/18
hinged back	13-16	18/18
inflammation of eyes/conjunctivitis	14-16	2/18

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# Infection of different organs & brain

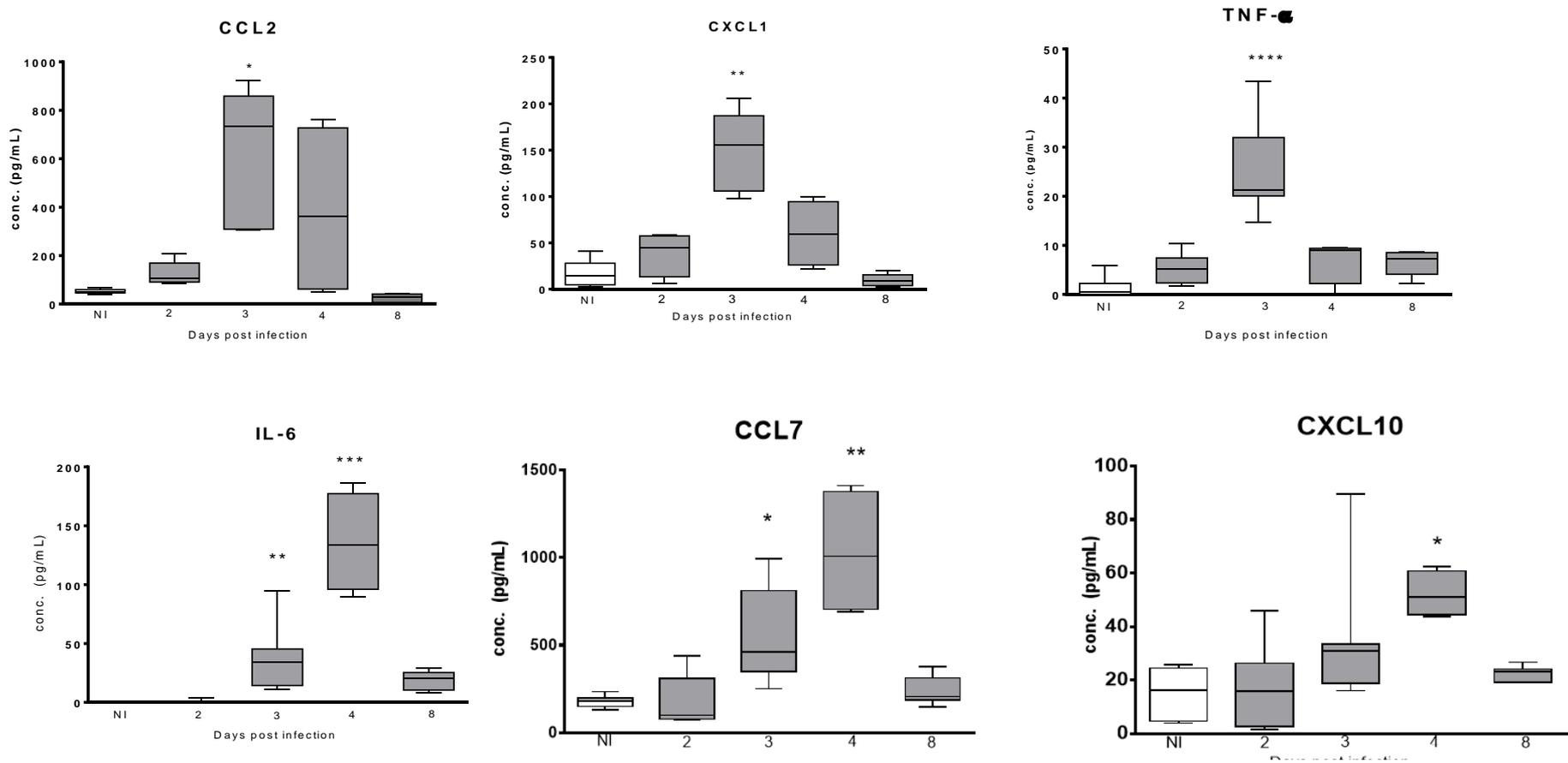


# Expression of pro-inflammatory cytokines and chemokines induced by ZIKV infection



In particular, **IFN- $\gamma$**  and **IL-18** were increased systematically during the course of infection.

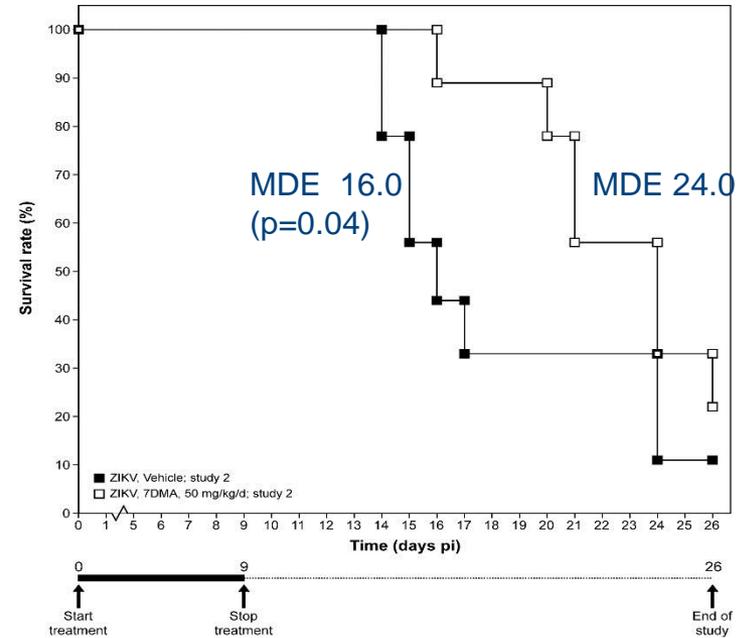
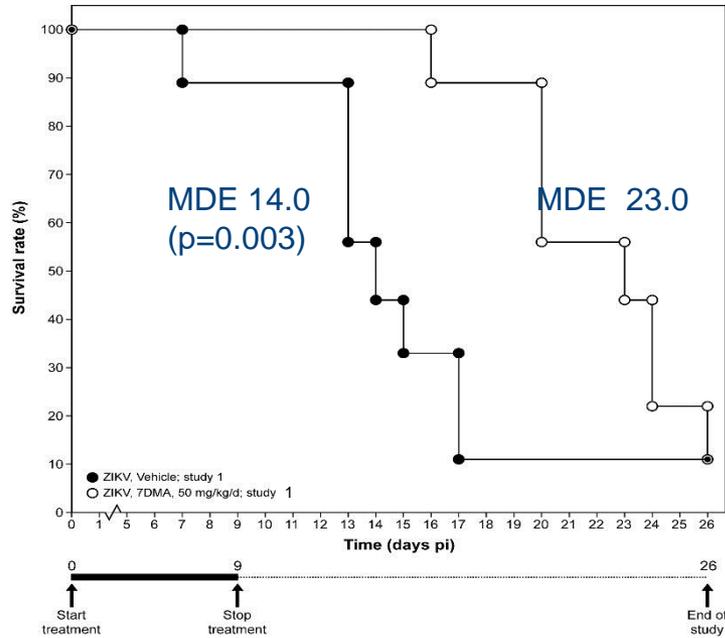
# Levels of pro-inflammatory cytokines and chemokines



Infection with ZIKV resulted in the induction of pro-inflammatory cytokines (IFN- $\gamma$ , IL18, TNF- $\alpha$ , IL-6) and chemokines (CCL2, CCL5, CCL7, CXCL1, CXCL10) in serum of AG129 mice.

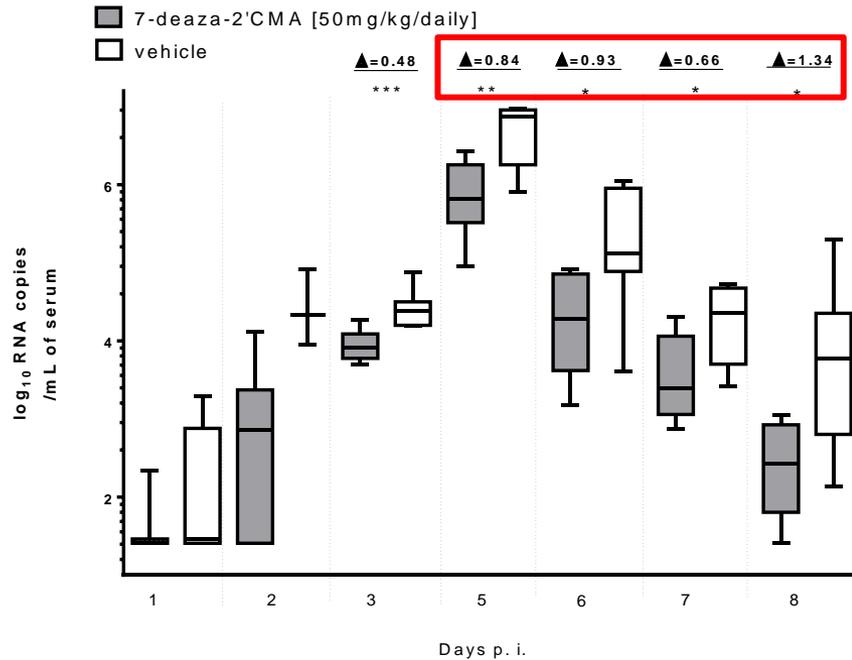
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# 7DMA delays ZIKV induced disease

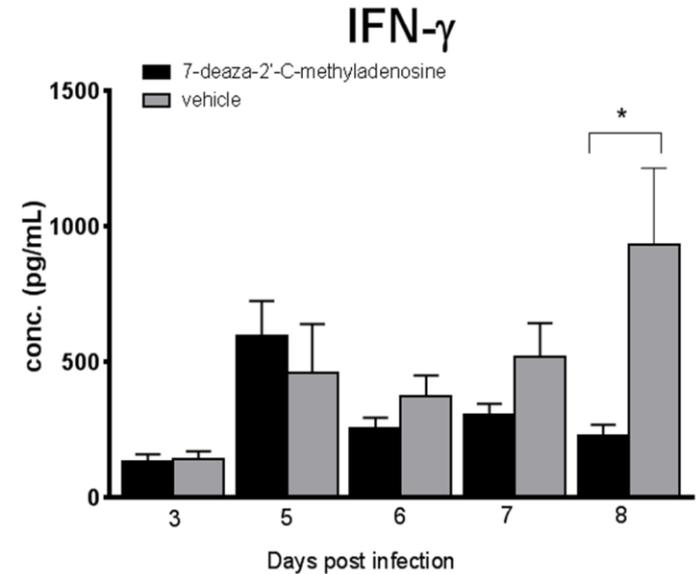


7DMA (50mg/kg/day, QD, for 10 days)

# 7DMA reduces viremia



Significant reduction of viral RNA load detected in serum:  $\sim 1\log_{10}$  reduction at d5-d7 pi



Significant reduction in serum levels of IFN- $\gamma$  at d8 pi



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beta  
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New Results

**The viral polymerase inhibitor 7-deaza-2'-C-methyladenosine is a potent inhibitor of in vitro Zika virus replication and delays disease progression in a robust mouse infection model**

Joanna Zmurko, Rafael E Marques, Dominique Schols, Erik Verbeken, Suzanne J.F. Kaptein, Johan Neyts

doi: <http://dx.doi.org/10.1101/041905>

This article is a preprint and has not been peer-reviewed [what does this mean?].

# The YFV vaccine is safe and highly effective but thermostable and there is a shortage



Max Theiler  
Nobelprize

Subculturing YFV Asibi strain >200 times in mice and cell culture: YFV-17D strain (1937)



Vaccination with  $>10^6$  pfu of YFV-17D: >99.5% protection



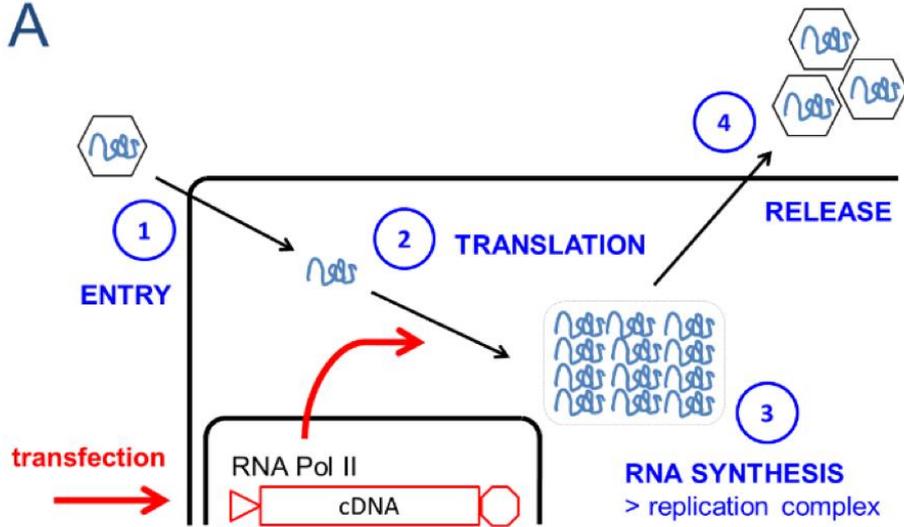
Sanofi Pasteur



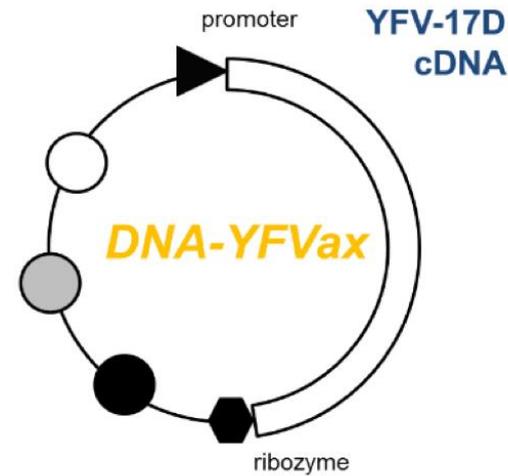
Bio-Manguinhos, Brasil

# DNA-YFVax technology: Replication of YFV-17D can be efficiently launched from plasmid DNA

A

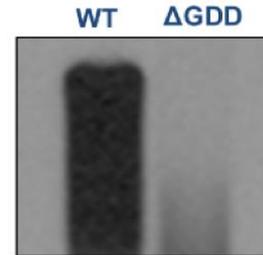


B

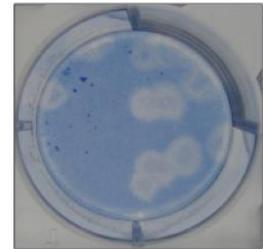


YFV-17D cloned in a BAC

C

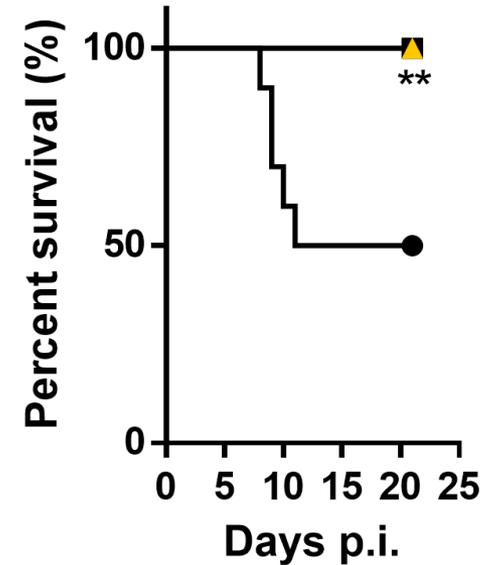
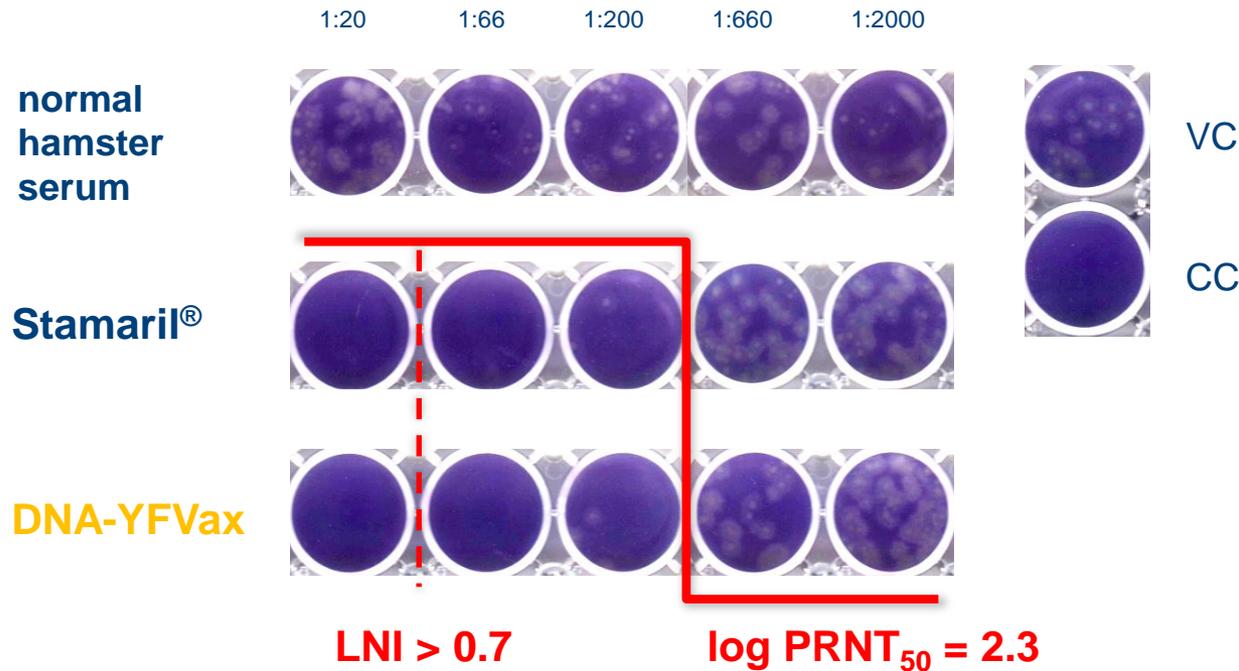


D



$\Delta$ GDD – replication deficient mutant with lethal deletion in the viral polymerase gene

# DNA-YFVax as effective as the commercial vaccine

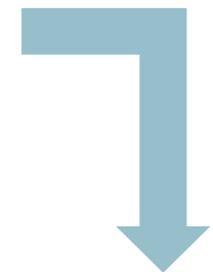
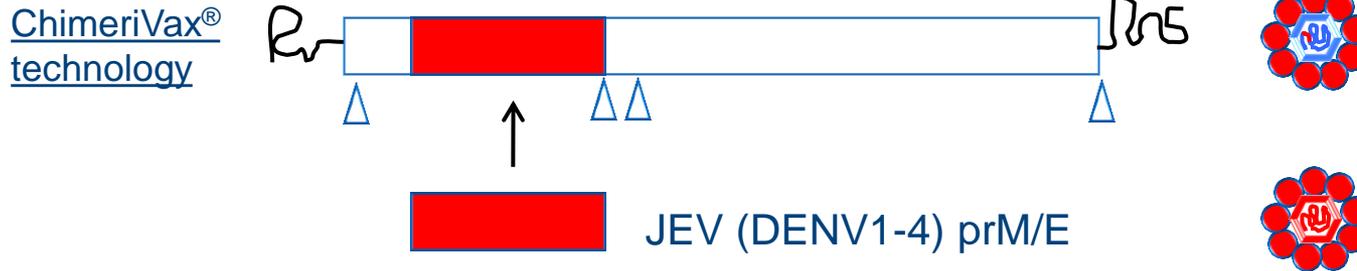
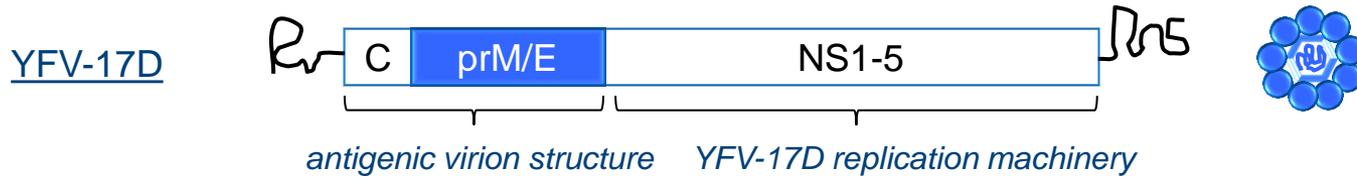


Plaque reduction neutralization test (PRNT) titers exceed log neutralization indices (LNI) that are considered immune correlates of protection\* by > 40-fold.

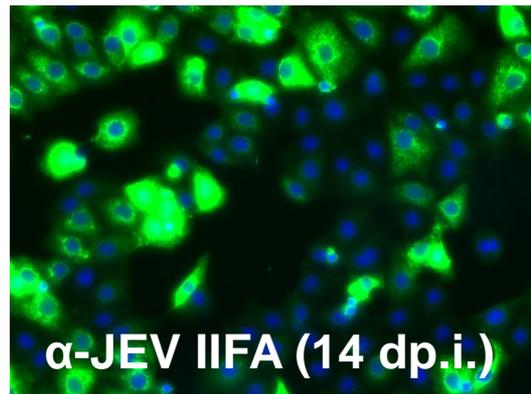
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\* WHO Wkly Epidemiol Rec 2013

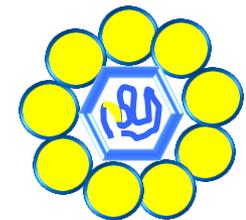
# DNA-YFVax approach can be used for other flavivirus vaccines viruses



POC:



Ongoing:



**DNA-ZIKVax**

# Conclusion

Assays available for HTS (in 384well plate format) and validation

AG129 ZIKV model validated for antiviral studies

- whether sexual transmission occurs
- study effect of infection on pubs

AG129 ZIKV model can likely also be used to initially assess vaccine efficacy

**DNA-YFVax** technology may allow to easily produce heat stable flavivirus vaccines as efficient as the parent vaccines

Pan-flavivirus drugs are needed that can be used for the treatment/prophylaxis of dengue, zika, JEV....

# Acknowledgements

## **Neyts-lab Leuven**

Joanna Zmurko

Rafael Elias

Carolien Dekeyser

Leen Delang

Suzanne Kaptein

Rafael Elias

Niraj Mishra

Dieudonné Buh Kum

Kai Dallmeier

## **Univ. Logan, Utah**

Justin Julander

## **NIAID**

Heather Greenstone

Christina Cassetti

## **Stem cell Institute Leuven**

Nicky Helsen

Tine Tricot,

Catherine Verfaillie